



German Journal of Veterinary Research

eISSN:2703-1322



Mini-review

Developing probiotics, prebiotics, and organic acids to control Salmonella spp. in commercial turkeys at the University of Arkansas, USA

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This article is published in the special issue: Turkey Diseases, Production and Management.





Article History:

Received: 04-May-2021 Accepted: 18-May-2021 *Corresponding author:

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Abstract

In the United States, non-typhoidal Salmonella causes over one million foodborne infections every year, and turkey meat contaminated with Salmonella has been associated from the farm to the processing plant. These outbreaks emphasize efforts to decrease and prevent human illness associated with live poultry contact through comprehensive interventions from "farm-to-fork" levels. This review article revises the role of the turkey upper respiratory tract, which is now known to play a crucial role in colonization and as a source of contamination, for this remarkable bacterium that has co-evolved to infect plants and animals. Because agriculture represents over 60% of the economy of the state of Arkansas, the mission of our laboratory over the last 21 years has been directed to evaluate and develop applied research to help reduce the incidence of Salmonella spp. from commercial turkey operations. A summary of the published research is presented.

Keywords: Salmonellosis, Turkeys, Ground meat, Nutraceuticals, Vaccines

Citation: Tellez-Isaias, G., Vuong, C.N., Graham, B.D., Selby, C.M., Graham, L.E., Sen~as-Cuesta, R., Barros, T.L., Beer, L.C., Coles, M.E., Forga, A.J., Ruff, J., Hernandez-Velasco, X. and Hargis, B.M. 2021. Developing probiotics, prebiotics, and organic acids to control Salmonella spp. in commercial turkeys at the University of Arkansas, USA. Ger. J. Vet. Res. 1(3): 7-12 https://doi.org/10.51585/gjvr.2021.3.0014

Introduction

Foodborne or waterborne microbial pathogens are associated with diarrheal disorders, killing an estimated two million people annually (Schlundt et al., 2004). In the United States, non-typhoidal Salmonella causes over one million foodborne infections every year (Scallan et al., 2011). Poultry meat, particularly ground turkey contaminated with Salmonella, has been associated from the farm to the processing plant (Ar- senault et al., 2007; Loharikar et al., 2012). These outbreaks emphasize the necessity and efforts to decrease and prevent human illness associated with live poultry contact through comprehensive interventions from "farm-to-fork" levels.

In current turkey processing, the organ or anatomical source of Salmonella contamination in products, especially ground turkey, is largely unreported. Hence, in a recent study, we evaluated the fate and organ distribution of Salmonella enterica serovar Reading in tur-

eys for up to three weeks, confirming that Salmonella Reading is an emerging problem for the turkey industry (Ashcraft et al., 2021). The first report of Salmonella Reading in commercial turkeys in the USA was published in 1956 (Mitrovic, 1956). Even though Salmonella Reading is a serotype that is uncommonly associated with human illness, during 2018-2019, four recalls of turkey meat were declared in 42 states in the USA reported three hundred and fifty-six Salmonella Reading linked ground turkey meat (Hassan et al., 2019). In combination with previous works completed by our laboratory, the results of this work indicate that the turkey upper respiratory tract (Figure 1) may play a much more significant role in Salmonella contamination during processing than previously known (Kallapura et al., 2014a). Laboratory and commercial trials have confirmed the respiratory route as a critical route for Salmonella spp. infection in chickens and turkeys (Kallapura et al., 2014b,c).

Positive tracheas



Figure 1: Evaluation of *Salmonella* spp. prevalence in trachea samples from commercially processed turkeys (Kallapura et al., 2014a). Trachea and ceca were aseptically removed from 16-week-old commercial turkeys at the processing plant (n=100). After aseptic removal from the birds, tracheas were clamped on each end, and 20 mL of peptone water was added for 8 hours of incubation at 37°C. The peptone water from each trachea was collected and enriched with an equal volume of 2X tetrathionate broth for overnight incubation. Samples were then streaked onto XLD-4 agar.

Salmonella incidence, virulence, undercooking, food consumption behavior, and host resistance play important roles in this critical foodborne pathogen, as has been described by (Oscar, 2021). Through eating, direct contact, and via the environment, the human and the animal bacterial flora are in contact with each other. The foodborne route is probably the most important gateway for this contact. Most infections with enteric zoonotic bacterial pathogens, such as Salmonella enterica occur through this route. Hence, food becomes an important essential vehicle for this zoonotic pathogen. In recent years, one of the major issues in food safety has been the lack of cross-sectoral cooperation among the food production chain, food control, human health sector, restaurants, and school cafeterias. The term "farm-to-fork" is a social movement that promotes serving local food at restaurants and school cafeterias with attitudes about food safety, including the food production chain. Therefore, science and education programs are necessary to reduce this zoonotic pathogen at relevant points of the "farm-to-fork" food production chain.

Holistic control methods for salmonellosis in turkeys

While antibiotics have saved millions of human and animal lives and have been used for decades as growth promoters, several studies have revealed that turkeys are a critical reservoir for multidrug-resistance (MDR) Salmonella strains in several countries around the world, reinforcing the need to develop alternative

strategies to reduce the development of antimicrobial resistance (Poppe et al., 1995; Beier et al., 2011; Choudhary et al., 2018; Yeh et al., 2018). Turkeys and chickens are highly vulnerable to Salmonella infection; therefore, decreasing the incidence of Salmonella before processing is vital. Salmonella enterica serovars remain significant foodborne pathogens due to the remarkable ability of this bacteria to infect all forms of life (Hernández-Reyes and Schikora, 2013), as indicated in Figure 2. Moreover, scientific, consumer, and global commercial strategic positions force producers to find alternatives to antibiotic growth promoters.

Since 2017, Salmonella enterica serovar Heidelberg has been the most frequently reported Salmonella from turkey ground meat in the USA, and a substantial number of the isolates are resistant to several antimicrobial agents (Nair et al., 2017; Nisar et al., 2017). In 2011, the Centers for Disease Control and Prevention identified a multistate cluster of Salmonella Heidelberg infections and two multidrug-resistant isolates from raw ground turkey retail samples (Routh et al., 2015). The outbreak resulted in the largest recall of fresh ground meat in the USA, involving 36 million pounds of turkey meat (Bearson et al., 2017).

Clearly, *Salmonella* has established a commensal-like condition in livestock and poultry, contributing to the asymptomatic carrier status of the human food-borne pathogen in our animal food supply. This is a critical situation if we consider that the acquisition of resistance plasmids to ceftiofur from *Salmonella* isolates from animals, including poultry, have been reported. Hence, in recent years, this reality has become a public, scientific, and health concern,

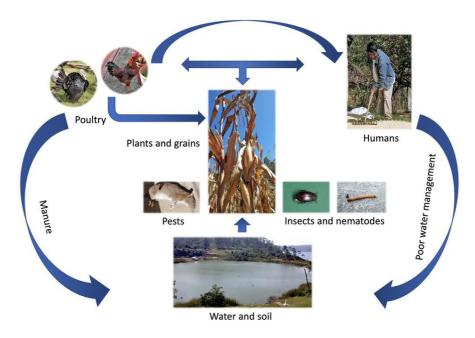


Figure 2: Salmonella cross-kingdom infection.

considering that third-generation cephalosporins are the antimicrobials of choice for the treatment of salmonellae infections in humans (Frye and Fedorka-Cray, 2007).

Figure 3 shows the infection routes of *Salmonella* in turkeys. Under this scenario, several control methods must be implemented to reduce the number of reproductive cases. Several studies have shown promising results using *Salmonella* spp. vaccines in turkeys (Zhang-Barber et al., 1999; Penha Filho et al., 2012; Hesse et al., 2017, 2018). The *Salmonella* Typhimurium BBS 866 vaccine strain has demonstrated cross-protection against diverse *Salmonella* serovars, not only in poultry but also in mammals (Hesse et al., 2016).

University of Arkansas research

Agriculture represents over 60% of the economy of the state of Arkansas, and poultry plays a critical role in this sector. As part of the Center of Excellence in Poultry Science, the John Kirkpatrick Skeeles Poultry Health Laboratory has the mission to listen to the demands and health problems of poultry companies and to develop applied research programs to solve those issues. As a result, over the last 21 years, our laboratory has developed and evaluated several probiotics, prebiotics, organic acids, and vaccines on commercial turkey farms with a high incidence of Salmonella spp. Those studies were part of the master's thesis and Ph.D. dissertations that were published at the University of Arkansas. More recently, we have also been evaluating phytogenic compounds. A summary of these experiences is described below.

In three commercial trials, the effects of selected probiotic bacteria or antibiotics on the performance of poults with idiopathic diarrhea and stunting were evaluated. Turkey poults that received antibiotics followed by a probiotic culture showed a significant improvement in body weight gain than non-treated probiotic control poults (Higgins et al., 2005).

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Following this trial, two probiotic cultures combined with a formulation of a commercial organic acid (OA) product were tested in turkey houses that had a historically high incidence of *Salmonella* spp. Two weeks after treatment, environmental *Salmonella* recovery was reduced by up to 100%, suggesting that the use of selected probiotic cultures in combination with OA is a reliable and effective method to reduce environmental *Salmonella* in turkey houses before processing (Jarquin et al., 2007; Vicente et al., 2007). These results were further confirmed in another field trial (Vicente et al., 2008).

The University of Arkansas then licensed the technology, and the lactic acid bacteria probiotic culture (FloraMax B-11) was tested alone or in combination with lactose as a prebiotic in commercial turkey houses during the brooding and grow-out phases. Results showed a remarkable increase in body weight gain dur- ing the starter phase that was maintained or enhanced during the grow-out phase (Torres-Rodriguez et al., 2007b). These results encouraged the turkey company to confirm and extend the trials to 118 commercial turkey houses, ranging from 1,542 to 30,390 hens per lot, with Nicholas or Hybrid genetic lines. The overall results indicated that the probiotic culture increased market body weight, showing a significant cost-benefit ratio (Torres-Rodriguez et al., 2007a).

In more recent studies with other lactic acid bacteria probiotic candidates, we have shown significant improvement in transepithelial electrical resistance *in vitro* and intestinal colonization of *Salmonella* Typhimurium in neonatal turkey poults (Yang et al., 2018) and *Salmonella* Enteritidis (Arreguin-Nava et al., 2019).

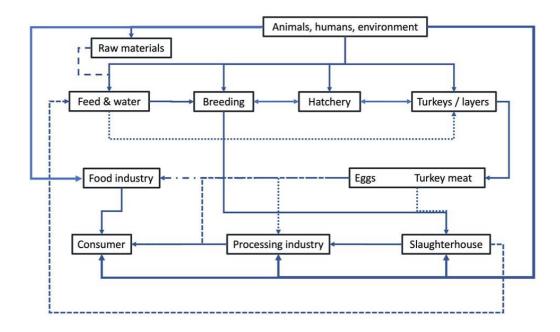


Figure 3: Infection routes of *Salmonella* in turkeys.

Phytogenic compounds

Phytogenic compounds and organic acids have also been extensively studied for their antioxidant, antiinflammatory, and immune-modulating properties against *Salmonella* spp. (Friedman et al., 2002, 2003, 2004; Bajpai et al., 2012). Our laboratory has also confirmed the benefits of these natural compounds, confirming that they are safe and serve as holistic alternatives to antibiotic growth promoters in poultry (Hernandez-Patlan et al., 2018a,b, 2019; Leyva-Diaz et al., 2021).

Vaccines

Our laboratory developed a recombinant oral vaccine using the conserved *fli* C that protects animals against multiple *Salmonella* serotypes. This vaccine was evaluated in three independent commercial turkey operations and has induced humoral response and reduction to *Salmonella* spp. infection (Wolfenden et al., 2010; Kremer et al., 2011).

Conclusions

No single "silver bullet" has been identified that can be ubiquitously applied commercially to eliminate Salmonella from commercial poultry. Success- ful poultry companies that have eliminated AGP and have been able to reduce the incidence of Salmonella from their operations have used several nutraceuticals and vaccines simultaneously in combination with strict biosecurity programs and genetic sources free of Salmonella spp. and Mycoplasma spp. with outstanding results and profitability.

Article Information

Funding. The research was supported in part by funds provided by USDA-NIFA Sustainable Agriculture Systems, Grant No. 2019-69012-29905. Title of Project: Empowering U.S. Broiler Production for Transformation and Sustainability USDA-NIFA (Sustainable Agriculture Systems): No. 2019-69012-29905.

Conflict of Interest. The authors declare no conflict of interest.

References

Arreguin-Nava, M.A., Hern'andez-Patl'an, D., Solis-Cruz, B., Latorre, J.D., Hernandez-Velasco, X., Tellez, G., El-Ashram, S., Hargis, B.M., Tellez-Isaias, G., 2019. Isolation and identification of lactic acid bacteria probiotic culture candidates for the treatment of *Salmonella enterica* serovar Enteritidis in neonatal turkey poults. Animals 9. 10.3390/ani9090696.

Arsenault, J., Letellier, A., Quessy, S., Normand, V., Boulianne, M., 2007. Prevalence and risk factors for *Salmonella* spp. and *Campylobacter* spp. caecal colonization in broiler chicken and turkey flocks slaughtered in Quebec, Canada. Preventive Veterinary Medicine 81, 250–264. 10.1016/j.prevetmed.2007. 04.016.

Ashcraft, A., Coles, M., Beer, L., Graham, B., Tellez-Isaias, G., Wooming, B., Hargis, B., 2021. Research note: Fate and dissemination of *Salmonella enterica* serovar Reading in turkeys at processing using an oral gavage challenge model. Poultry Science, 10111410.1016/j.psj.2021.101114.

Bajpai, V.K., Baek, K.H., Kang, S.C., 2012. Control of *Salmonella* in foods by using essential oils: A review. Food Research International 45, 722–734. 10.1016/j.foodres. 2011.04.052.

Bearson, B.L., Bearson, S.M.D., Looft, T., Cai, G., Shippy, D.C., 2017. Characterization of a multidrug-resistant *Salmonella* enterica serovar Heidelberg outbreak strain in commercial turkeys: Colonization, transmission, and host transcriptional response. Frontiers in Veterinary Science 4, 156. 10.3389/fvets.2017.00156.

Beier, R.C., Anderson, P.N., Hume, M.E., Poole, T.L., Duke, S.E., Crippen, T.L., Sheffield, C.L., Caldwell, D.J., Byrd,

- J.A., Anderson, R.C., Nisbet, D.J., 2011. Characterization of *Salmonella enterica* isolates from turkeys in commercial processing plants for resistance to antibiotics, disinfectants, and a growth promoter. Foodborne Pathogens and Disease 8, 593–600. 10.1089/fpd.2010.0702.
- Choudhary, M., Choudhary, B.K., Bhoyar, S., Kale, S.B., Chaudhari, S.P., Bera, B.C., Jain, A., Barbuddhe, S.B., 2018. Isolation and characterization of multidrug-resistant *Leclercia* species from animal clinical case. Letters in Applied Microbiology 66, 44–48. 10.1111/lam.12811.
- Friedman, M., Henika, P.R., Levin, C.E., Mandrell, R.E., 2004. Antibacterial activities of plant essential oils and their components against *Escherichia coli* O157:H7 and *Salmonella enterica* in apple juice. Journal of Agricultural and Food Chemistry 52, 6042–6048. 10.1021/jf0495340.
- Friedman, M., Henika, P.R., Mandrell, R.E., 2002. Bacteriadal activities of plant essential oils and some of their isolated constituents against *Campylobacter jejuni, Escherichia coli, Listeria monocytogenes*, and *Salmonella enterica*. Journal of Food Protection 65, 1545–1560. 10.4315/0362-028x-65.10. 1545.
- Friedman, M., Henika, P.R., Mandrell, R.E., 2003. Antibacterial activities of phenolic benzaldehydes and benzoic acids against *Campylobacter jejuni, Escherichia coli, Listeria monocytogenes*, and *Salmonella enterica*. Journal of Food Protection 66, 1811–1821. 10.4315/0362-028x-66.10.1811.
- Frye, J.G., Fedorka-Cray, P.J., 2007. Prevalence, distribution and characterization of ceftiofur resistance in *Salmonella enterica* isolated from animals in the USA from 1999 to 2003. International Journal of Antimicrobial Agents 30, 134–142. 10.1016/j.ijantimicag.2007.03.013.
- Hassan, R., Buuck, S., Noveroske, D., Medus, C., Sorenson, A.,
 Laurent, J., Rotstein, D., Schlater, L., Freiman, J., Douris,
 A., Simmons, M., Donovan, D., Henderson, J., Tewell, M.,
 Snyder, K., Oni, O., Von Stein, D., Dassie, K., Leeper, M.,
 Adediran, A., Dowell, N., Gieraltowski, L., Basler, C., 2019.
 Multistate outbreak of *Salmonella* infections linked to raw
 turkey products United States, 2017-2019. MMWR.
 Morbidity and Mortality Weekly Report 68, 1045-1049.
 10.15585/ mmwr6846a1.
- Hernandez-Patlan, D., Solis-Cruz, B., M'endez-Albores, A., Latorre, J.D., Hernandez-Velasco, X., Tellez, G., L'opez-Arellano, R., 2018a. Comparison of PrestoBlue® and plating method to evaluate antimicrobial activity of ascorbic acid, boric acid, and curcumin in an *in-vitro* gastrointestinal model. Journal of Applied Microbiology 124, 423–430. 10.1111/jam.13659.
- Hernandez-Patlan, D., Solis-Cruz, B., Pontin, K.P., Latorre, J.D., Baxter, M.F.A., Hernandez-Velasco, X., Merino-Guzman, R., M´endez-Albores, A., Hargis, B.M., Lopez-Arellano, R., Tellez, G., 2018b. Evaluation of a solid dispression of curcumin with polyvinylpyrrolidone and boric acid against *Salmonella* Enteritidis infection and intestinal permeability in broiler chickens: A pilot study. Frontiers in Microbiology 9, 1289. 10.3389/fmicb.2018.01289.
- Hernandez-Patlan, D., Sol´ıs-Cruz, B., Patrin Pontin, K., Latorre, J.D., Baxter, M.F.A., Hernandez-Velasco, X., Merino-Guzman, R., M´endez-Albores, A., Hargis, B.M., Lopez-Arellano, R., Tellez-Isaias, G., 2019. Evaluation of the dietary supplementation of a formulation containing

- ascorbic acid and a solid dispersion of curcumin with boric acid against *Salmonella* Enteritidis and necrotic enteritis in broiler chickens. Animals 9. 10.3390/ani9040184.
- Hern´andez-Reyes, C., Schikora, A., 2013. *Salmonella*, a cross-kingdom pathogen infecting humans and plants. FEMS Microbiology Letters 343, 1–7. 10.1111/1574-6968.12127.
- Hesse, M., Stamm, A., Berndt, A., Glu"nder, G., Weber, R., 2017. Immune response to *salmonella* infections in vaccinated and non-vaccinated turkeys. Research in Veterinary Science 115, 165–173. 10.1016/j.rvsc.2017.04.002.
- Hesse, M., Stamm, A., Weber, R., Glu"nder, G., 2018. Efficacy of a *Salmonella* live vaccine for turkeys in different age groups and antibody response of vaccinated and non-vaccinated turkeys. BMC Research Notes 11, 431. 10.1186/ s13104-018-3524-1.
- Hesse, M., Stamm, A., Weber, R., Glu"nder, G., Berndt, A., 2016. Immune response of turkey poults exposed at 1 day of age to either attenuated or wild *Salmonella* strains. Veterinary Immunology and Immunopathology 174, 1–10. 10.1016/j.vetimm.2016.04.004.
- Higgins, S., Torres-Rodriguez, A., Vicente, J., Sartor, C., Pixley, C., Nava, G., Tellez, G., Barton, J., Hargis, B., 2005. Evaluation of intervention strategies for idiopathic diarrhea in commercial turkey brooding houses. Journal of Applied Poultry Research 14, 345–348. 10.1093/japr/14.2.345.
- Jarquin, R., Nava, G., Wolfenden, A., Donoghue, A., Hanning, I., Higgins, S., Hargis, B., 2007. The evaluation of organic acids and probiotic cultures to reduce *Salmonella* Enteriditis horizontal transmission and crop infection in broiler chickens. International Journal of Poultry Science 6, 182–186. 10.3923/ijps.2007.182.186.
- Kallapura, G., Botero, A., Layton, S., Bielke, L., Latorre, J., Menconi, A., Hern´andez-Velasco, X., Bueno, D., Hargis, B., T´ellez, G., 2014a. Evaluation of recovery of *Salmonella* from trachea and ceca in commercial poultry. Journal of Applied Poultry Research 23, 132–136. 10.3382/japr.2013-00854.
- Kallapura, G., Kogut, M.H., Morgan, M.J., Pumford, N.R., Bielke, L.R., Wolfenden, A.D., Faulkner, O.B., Latorre, J.D., Menconi, A., Hernandez-Velasco, X., Kuttappan, V.A., Hargis, B.M., Tellez, G., 2014b. Fate of Salmonella Senftenberg in broiler chickens evaluated by challenge experiments. Avian Pathology 43, 305–309. 10.1080/03079457.2014.923554.
- Kallapura, G., Morgan, M.J., Pumford, N.R., Bielke, L.R., Wolfenden, A.D., Faulkner, O.B., Latorre, J.D., Menconi, A., Hernandez-Velasco, X., Kuttappan, V.A., Hargis, B.M., Tellez, G., 2014c. Evaluation of the respiratory route as a viable portal of entry for *Salmonella* in poultry via intratracheal challenge of *Salmonella* Enteritidis and *Salmonella* Typhimurium. Poultry Science 93, 340–346. 10.3382/ps.2013-03602.
- Kremer, C.J., O'Meara, K.M., Layton, S.L., Hargis, B.M., Cole, K., 2011. Evaluation of recombinant *Salmonella* expressing the flagellar protein fliC for persistence and enhanced antibody response in commercial turkeys. Poultry Science 90, 752–758. 10.3382/ps.2010-01076.
- Leyva-Diaz, A.A., Hernandez-Patlan, D., Solis-Cruz, B., Adhikari, B., Kwon, Y.M., Latorre, J.D., Hernandez-Velasco, X., Fuente-Martinez, B., Hargis, B.M., Lopez-Arellano, R.,

- Tellez-Isaias, G., 2021. Evaluation of curcumin and copper acetate against *Salmonella* Typhimurium infection, intestinal permeability, and cecal microbiota composition in broiler chickens. Journal of Animal Science and Biotechnology 12, 23. 10.1186/s40104-021-00545-7.
- Loharikar, A., Briere, E., Schwensohn, C., Weninger, S., Wagendorf, J., Scheftel, J., Garvey, A., Warren, K., Villamil, E., Rudroff, J.A., Kurkjian, K., Levine, S., Colby, K., Morrison, B., May, A., Anderson, S., Daly, E., Marsden-Haug, N., Erdman, M.M., Gomez, T., Rhorer, A., Castleman, J.,Adams, J.K., Theobald, L., Lafon, P., Trees, E., Mitchell, J., Sotir, M.J., Behravesh, C.B., 2012. Four multistate outbreaks of human *Salmonella* infections associated with live poultry contact, United States, 2009. Zoonoses and Public Health 59, 347–354. 10.1111/j.1863-2378.2012.01461.x.
- Mitrovic, M., 1956. First report of paratyphoid infection in turkey poults due to *Salmonella* Reading. Poultry Science 35, 171–174. 10.3382/ps.0350171.
- Nair, D.V.T., Vazhakkattu Thomas, J., Noll, S., Porter, R., Kollanoor Johny, A., 2017. Effect of various inoculum levels of multidrug-resistant Salmonella enterica serovar Heidelberg (2011 ground turkey outbreak isolate) on cecal colonization, dissemination to internal organs, and deposition in skeletal muscles of commercial turkeys after experimental oral challenge. Frontiers in Microbiology 8, 2680. 10.3389/fmicb. 2017.02680.
- Nisar, M., Kassem, I.I., Rajashekara, G., Goyal, S.M., Lauer, D., Voss, S., Nagaraja, K.V., 2017. Genotypic relatedness and antimicrobial resistance of *Salmonella* Heidelberg isolated from chickens and turkeys in the Midwestern United States. Journal of Veterinary Diagnostic Investigation 29, 370–375. 10.1177/1040638717690784.
- Oscar, T., 2021. *Salmonella* prevalence alone is not a good indicator of poultry food safety. Risk Analysis 41, 110–130. 10.1111/risa.13563.
- Penha Filho, R.A.C., Moura, B.S., de Almeida, A.M., Montassier, H.J., Barrow, P.A., Berchieri Junior, A., 2012. Hu- moral and cellular immune response generated by different vaccine programs before and after *Salmonella* Enteritidis challenge in chickens. Vaccine 30, 7637–7643. 10.1016/j.vaccine.2012.10.020.
- Poppe, C., Kolar, J.J., Demczuk, W.H., Harris, J.E., 1995. Drug resistance and biochemical characteristics of *Salmonella* from turkeys. Canadian Journal of Veterinary Research 59, 241–248. URL: https://www.ncbi.nlm.nih.gov/pubmed/8548684.
- Routh, J.A., Pringle, J., Mohr, M., Bidol, S., Arends, K., Adams-Cameron, M., Hancock, W.T., Kissler, B., Rickert, R., Folster, J., Tolar, B., Bosch, S., Barton Behravesh, C., Williams, I.T., Gieraltowski, L., 2015. Nationwide outbreak of multidrug-resistant *Salmonella* Heidelberg infections associated with ground turkey: United States, 2011. Epidemiology and Infection 143, 3227–3234. 10.1017/S0950268815000497.

- Scallan, E., Hoekstra, R., Angulo, F., Tauxe, R., Widdowson, M., Roy, S., Jones, J., Griffin, P., 2011. Foodborne illness acquired in the United States pathogens. Emerging Infectious Diseases 17, 7–15. 10.3201/eid1701.091101p1.
- Schlundt, J., Toyofuku, H., Jansen, J., Herbst, S.A., 2004. Emerging food-borne zoonoses. Revue Scientifique et Technique (International Office of Epizootics) 23, 513– 533. 10. 20506/rst.23.2.1506.
- Torres-Rodriguez, A., Donoghue, A.M., Donoghue, D.J., Barton, J.T., Tellez, G., Hargis, B.M., 2007a. Performance and condemnation rate analysis of commercial turkey flocks treated with a *Lactobacillus* spp.-based probiotic. Poultry Science 86, 444–446. 10.1093/ps/86.3.444.
- Torres-Rodriguez, A., Higgins, S., Vicente, J., Wolfenden, A., Gaona-Ramirez, G., Barton, J., Tellez, G., Donoghue, A., Hargis, B., 2007b. Effect of lactose as a prebiotic on turkey body weight under commercial conditions. Journal of Applied Poultry Research 16, 635–641. 10.3382/japr.2006-00127.
- Vicente, J., Higgins, S., Bielke, L., Tellez, G., Donoghue, D., Donoghue, A., Hargis, B., 2007. Effect of probiotic culture candidates on *Salmonella* prevalence in commercial turkey houses. Journal of Applied Poultry Research 16, 471–476. 10.1093/japr/16.3.471.
- Vicente, J.L., Torres-Rodriguez, A., Higgins, S.E., Pixley, C., Tellez, G., Donoghue, A.M., Hargis, B.M., 2008. Effect of a selected *Lactobacillus* spp.-based probiotic on *Salmonella enterica* serovar Enteritidis infected broiler chicks. Avian Diseases 52, 143–146. 10.1637/7847-011107-ResNote.
- Wolfenden, R.E., Layton, S.L., Wolfenden, A.D., Khatiwara, A., Gaona-Ram´ırez, G., Pumford, N.R., Cole, K., Kwon, Y.M., Tellez, G., Bergman, L.R., Hargis, B.M., 2010. Development and evaluation of candidate recombinant *Salmonella* vectored *Salmonella* vaccines. Poultry Science 89, 2370–2379. 10.3382/ps.2010-00702.
- Yang, Y., Latorre, J.D., Khatri, B., Kwon, Y.M., Kong, B.W., Teague, K.D., Graham, L.E., Wolfenden, A.D., Mahaffey, B.D., Baxter, M., Hernandez-Velasco, X., Merino-Guzman, R., Hargis, B.M., Tellez, G., 2018. Characterization and evaluation of lactic acid bacteria candidates for intestinal epithelial permeability and Salmonella Typhimurium colonization in neonatal turkey poults. Poultry Science 97, 515–521. 10.3382/ps/pex311.
- Yeh, J.C., Chen, C.L., Chiou, C.S., Lo, D.Y., Cheng, J.C., Kuo, H.C., 2018. Comparison of prevalence, phenotype, and antimicrobial resistance of *Salmonella* serovars isolated from turkeys in Taiwan. Poultry Science 97, 279–288. 10.3382/ps/pex293.
- Zhang-Barber, L., Turner, A., Barrow, P., 1999. Vaccination for control of *Salmonella* in poultry. Vaccine 17, 2538–2545. 10.1016/S0264-410X(99)00060-2.